

Please read this package insert carefully before use.

# Capilia™ Adeno Neo

## INTENDED USE

To detect adenovirus antigens in pharyngeal swab, nasal swab, nasal aspirate or keratoconjunctivitis swab (to assist in the diagnostic of adenovirus infectious disease).

## SUMMARY AND EXPLANATION OF THE TEST

Adenovirus is responsible for 5–10% of upper respiratory infections in children, and many infections in adults as well.

Common symptoms are chills, fever, sore throat, coughing, pinkeye, gastroenteritis and hemorrhagic cystitis.

Diagnosis is difficult because the initial symptoms can be similar to those caused by other infectious agents.

There are no antiviral drugs to treat adenoviral infections, so treatment is largely directed at the symptoms.

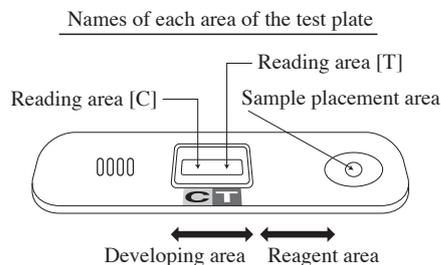
Because the adenovirus is highly contagious, rapid diagnosis and prompt treatment of patients can have a positive effect on public health.

Capilia Adeno Neo can provide rapid and accurate detection of adenovirus antigens from symptomatic patients. No special instruments or equipment are required.

## PRINCIPLE OF THE TEST

Measurement using this product is based on an immunochromatography assay using a monoclonal antibody that recognizes adenovirus antigens.

This product comprises a test plate with a carrier strip containing a sample placement area, a reagent area including a colloidal platinum-gold labeled anti-adenovirus monoclonal antibody (mouse) (hereinafter referred to as “colloidal platinum-gold labeled anti-adenovirus antibody”), a reading area [T] that fixes the anti-adenovirus monoclonal antibody (mouse) (hereinafter referred to as “anti-adenovirus antibody”), and a reading area [C] that fixes an anti-mouse immunoglobulin polyclonal antibody (rabbit) (hereinafter referred to as “anti-mouse immunoglobulin antibody”).



When a sample is placed on the sample placement area of the test plate, the colloidal platinum-gold labeled anti-adenovirus antibody dissolves and forms an immune complex with the adenovirus antigens in the sample. This immune complex migrates through the developing area by capillary action, is captured by the anti-adenovirus antibody fixed in the developing area, and forms a black line of colloidal platinum-gold in the reading area [T]. The black line visually displays the existence of adenovirus antigens in the sample.

Regardless of the existence of adenovirus antigens in the sample, excess colloidal platinum-gold labeled anti-adenovirus antibodies further migrate through the developing area, are captured by anti-mouse immunoglobulin antibodies fixed in the reading area, and form a black line in the reading area [C]. This means the colloidal platinum-gold labeled anti-adenovirus antibodies have migrated normally.

## REAGENTS AND MATERIALS PROVIDED

**[REF]** CAAD0370 Capilia Adeno Neo (20 Tests)

Test plates

- Components
  - Colloidal platinum-gold labeled anti-adenovirus monoclonal antibody (mouse)
  - Anti-adenovirus monoclonal antibody (mouse)

Extraction Buffer (to be used equally with the four products.) <sup>Note</sup>

- Components
  - Buffer, detergent, sodium azide (0.09%)

<sup>Note</sup> The extraction buffer is able to be used equally with the four products below:

- Capilia Flu Neo (rapid test for detecting Influenza virus antigen)
- Capilia Adeno Neo (rapid test for detecting Adenovirus antigen)
- Capilia RSV Neo (rapid test for detecting RS virus antigen)
- Capilia hMPV (rapid test for detecting human metapneumovirus antigen)

Specimen	Flu	Adeno	RSV	hMPV
Nasal swab	○	○	○	○
Nasal aspirate	○	○	○	○
Nasal discharge/Nasal mucus	○	×	×	×
Pharyngeal swab	○	○	×	○
Keratoconjunctivitis swab	×	○	×	×

Nozzles

**[REF]** CAAD0371 Capilia Adeno Neo Test Plate (10 Tests)

Test plates

- Components
  - Colloidal platinum-gold labeled anti-adenovirus monoclonal antibody (mouse)
  - Anti-adenovirus monoclonal antibody (mouse)

## MATERIALS REQUIRED BUT NOT PROVIDED

Timer, micropipette, pipette tips, suction machine, suction trap, sterile swabs (as listed here)

## Recommended Swab

The following swabs are recommended for use with the kit.

### For Nasal Swab

- ① FLOQSwabs™ (Cat No. 534CS01, Copan Italia S.p.A, Italy)
- ② Sterilized Swab P156A 10 pcs (Cat No. 4124, HEIWA MEDIC. CO., LTD, Japan)

### For Pharyngeal Swab

- ① FLOQSwabs™ (Cat No. 502CS01, Copan Italia S.p.A, Italy)
- ② Sterilized Swab PL6S 10 pcs (Cat No. 4371, HEIWA MEDIC. CO., LTD, Japan)

To collect nasal aspirate, any of the above swabs may be used.

## Acceptable Swab

The following swabs are acceptable for use with the kit.

- Tip material
  - Rayon, flocked nylon and polyester
- Standard tip size
  - For nasal swab**
  - Plain dry swab: maximum diameter 3 mm, length 12 mm
  - Flocked swab: maximum diameter 3 mm, length 15 mm
  - For pharyngeal swab**
  - Plain dry swab: maximum diameter 6 mm, length 14 mm
  - Flocked swab: maximum diameter 6 mm, length 16 mm
- Shaft material
  - Paper, plastic (PS, nylon), aluminum

## Unacceptable Swab

Do not use calcium alginate swabs.

## WARNING AND PRECAUTIONS

### 1. Precautions when handling (including hazard control)

- 1) Handle all the specimens as if they contain infectious agents.
- 2) In consideration of the risk of infection, wear protective clothes such as a mask and gloves and handle the specimens and samples carefully during the test.
- 3) If the extraction buffer gets into your eyes, immediately flush with a large quantity of water for 15 minutes or more. If you still feel some abnormality, see a doctor for treatment.
- 4) If the extraction buffer comes into contact with your hands or clothes, wash your hands and/or clothes with soap and a large quantity of water.

### 2. Precautions when using

- 1) This product is a rapid test for detecting adenovirus antigen.  
**A definite diagnosis should be made by an attending physician, in combination with the clinical symptoms, the result of a virus isolation culture test and other test results.**
- 2) Avoid touching saliva when collecting pharyngeal swab.
- 3) This product should be used in accordance with the procedure stated in the package insert.
- 4) In order to prevent deterioration, this product should be stored between 2°C and 30°C, avoiding high temperatures, high humidity and direct sunlight.
- 5) If this product has been refrigerated, it must be removed from the refrigerator at least 30 minutes before to be acclimatized to room temperature.
- 6) **The aluminum pouch containing a test plate should not be opened until the test plate is about to be used.**
- 7) The sample placement area and the reading area of the test plate should not be touched with the hands.
- 8) A precipitate may be seen in the extraction buffer, but the product can be used as it is, because the precipitate has been shown not to affect test results.
- 9) Do not use a swab if it is broken, bent or stained.
- 10) For nasal sampling, do not keep forcibly insert the swab, when the distance to the site is clearly shorter than usual. In particular, there is the possibility of resistance being imposed on the stick when the sample is collected from an infant or a patient with a narrow nasal cavity. In such a case, do not swab hard, exerting force on the stick. Moreover, do not rotate the stick forcibly.
- 11) For nasal sampling, any mass of mucus on the tip of the swab should be gently removed with gauze. Do not wipe the tip too hard. Mucosal epidermal cells should remain on the tip for testing.
- 12) Do not use any products beyond the expiration date.

### 3. Precautions for disposal

- 1) Because used test plates, swabs, tubes and nozzles after use, remaining samples, etc. may cause infections, they should be autoclaved (121°C, 20 min) or soaked in 0.1% sodium hypochlorite for more than one hour. When reagents, remaining reagents or their accessories are disposed of, they should be treated in accordance with the laws and regulations concerning medical waste disposal and water pollution control.
- 2) In the extraction buffer, 0.09% of sodium azide is included as a preservative. When solutions containing sodium azide continue to be discarded over a long period of time, explosive metallic azide may be produced if a drain is made of metal. Therefore, they should be discarded with a large quantity of water.

## STORAGE CONDITIONS

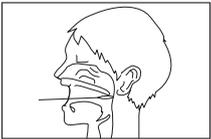
Storage : Store at 2°C to 30°C. **DO NOT FREEZE.**

Keep away from direct sunlight.

Do not use test plate or extraction buffer after expiration date.

## SPECIMEN COLLECTION AND PREPARATION

### 1. Methods of specimen collection



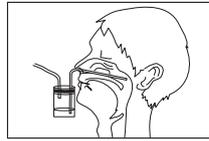
#### 1) Sampling of pharyngeal swab

Firmly insert a pharyngeal swab into the pharynx through the oral cavity, and collect the mucosal epithelium by swabbing the posterior wall of the pharynx and the palatine tonsil several times, centering around the rubefacient portion. Avoid touching saliva. If the specimen is mixed with saliva, the test result lines may become fainter on the test plate.



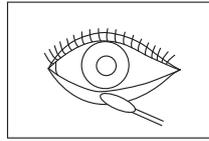
#### 2) Sampling of nasal swab

Firmly insert a nasal swab into the nasal cavity and collect mucosal epithelium by swabbing the nasal turbinate several times.



### 3) Sampling of nasal aspirate

Firmly insert one tube of the suction trap into the suction pump, and the other tube into a nasal cavity through an external nostril. Collect the nasal discharge aspirate in the suction trap by operating the suction pump. Soak a swab in the nasal aspirate collected by the trap, and let the swab absorb the nasal aspirate well. When nasal aspirate is taken using a micropipette or other instruments, dilute the nasal aspirate twofold with physiological saline and sample 200 µL of this dilution.

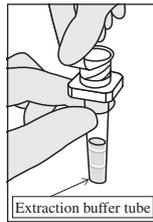


### 4) Sampling of keratoconjunctivitis swab

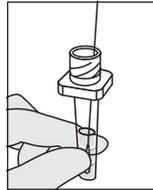
Use a pharyngeal swab, and collect the cuticle by scratching hard the keratoconjunctivitis several times. If necessary, use a surface anesthetic and scratch the area of inflammation as strongly as you can.

**With a collected specimen, prepare the sample according to the procedure specified in the following "Sample preparation" section and perform the test as quickly as possible.**

## 2. Sample preparation



Remove the aluminum sealing cap from the extraction buffer tube, while taking care not to spill the liquid.

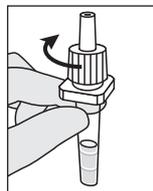


Soak the swab that collected the specimen in the extraction buffer, and stir well.

Then, pinch the tip of the swab firmly with the soft wall of the extraction buffer tube with your fingers and squeeze out the swab. Use this squeezed-out liquid as the sample.

When using a nasal aspirate specimen diluted twofold with physiological saline, add 200 µL of the specimen to the extraction buffer in the tube, and mix well. Use this mixture as the sample.

## TEST PROCEDURE



- 1) Firmly attach the nozzle (with a filter) provided in the kit to the top of the extraction buffer tube.
- 2) Hold the middle of the tube with the fingers and dispense **3 drops of the sample (80–120 µL)** onto the sample placement area of the test plate. Hold the tube perpendicularly and take care not to let the tip of the nozzle touch the sample placement area.
- 3) Observe the reading area of the test plate after **3 to 5 minutes** and interpret the result according to the "READING TEST RESULTS."

## READING TEST RESULTS

Allow the samples to react according to the procedure and read the black lines that appear in the reading area.

#### Positive



When black lines are seen at both [T] and [C] in the reading area (two lines), the result is read as positive. When a very faint black line is seen in the reading area [T], the result is also interpreted as positive.

#### Negative



When no black line is seen at [T] in the reading area but a black line is seen only at [C] in the reading area (one line), the result is read as negative.

When a black line at [C] in the reading area is faint but visually recognizable, chromatographic development has occurred normally.

#### Retesting



When no black line is seen at [C] in the reading area, there may be some problem with the test procedure or the reagent quality. The test should be performed again, using another test plate.



**A line that appears anywhere within the sections of the reading area, which are separated by color, is considered valid.**

**(Note)**

- When black lines are seen at both [T] and [C] in the reading area 3 to 5 minutes after dispensing the sample, the result is read as positive. No black line at [T] in the reading area even 5 minutes after dispensing the sample indicates a negative result.
- Do not use the test plate for reading a result beyond the judgment time as the result may change due to drying, etc.  
If the amount of antigen in the sample is very low and close to the detection limit of this product, a black line may appear at [T] in the reading area after 5 minutes or more of the judgment time, due to the feature of immunochromatography. In addition, due to a non-specific reaction caused by specimen-derived components, a black line may appear at [T] in the reading area after 5 minutes or more of the judgment time in rare cases.  
A definite diagnosis should be made comprehensively, not only based on the test result of this product but also taking other test results and clinical symptoms into account.
- A black line may not appear at [C] in the reading area due to problems with the test procedure or the reagent quality. In this case, the test should be performed again, using another test plate. If the same result is obtained in the re-test, try the test once more using the sample diluted twofold with saline as the black line may not appear at [C] in the reading area due to a factor in the specimen or the effect of saliva.
- If the amount of antigen is very high, a very thick line may be seen at [T] in the reading area and no black line may be seen at [C] in the reading area. In that case, dilute the sample with more extraction buffer and perform the test again.  
Example) Method for dilution of sample : Dispense 3 drops of the sample to a new extraction buffer tube, mix thoroughly and use the solution as the test sample.
- The line is valid even if there is unevenness in depth and there are breaks in the line.

**LIMITATIONS**

- This product is a rapid test for detecting adenovirus antigen.  
A definite diagnosis should be made by an attending physician, in combination with the clinical symptoms, the result of virus isolation culture test and other test results.
- An adenovirus is a highly contagious virus. Take appropriate steps to prevent in-hospital infection.
- When collecting the pharyngeal swab, avoid touching saliva. If the specimen is mixed with saliva, the test result lines may become fainter on the test plate.
- The test plate should be used immediately after opening the packaging. When it absorbs moisture, the quality deteriorates and an accurate result cannot be obtained.
- This product should be used for *in vitro* diagnosis only and should not be used for any other purposes.
- Please use this product following the operational method described in this package insert. We cannot guarantee results obtained from any other operations and for any other purposes that are not described in the package insert.
- The extraction buffer contains sodium azide. If the solution comes into contact with eye or mouth or adheres to the skin by mistake, take emergency measures such as thorough washing with water and seek medical treatment, if necessary.

**PERFORMANCE CHARACTERISTICS**

**1. Clinical data**

The result of the clinical performance evaluation in Japan (Comparison with approved products)

Kind of samples	Sensitivity (%)	Specificity (%)	Accuracy (%)	Total number
Pharyngeal swab	100 (56/56)	99.2 (125/126)	99.5 (181/182)	182

The result of domestic clinical performance evaluation (Comparison with PCR)

Kind of samples	Sensitivity (%)	Specificity (%)	Accuracy (%)	Total number
Nasal swab	96.5 (55/57)	98.2 (56/57)	97.4 (111/114)	114
Nasal aspirate	98.1 (52/53)	98.3 (58/59)	98.2 (110/112)	112
Keratoconjunctivitis swab	91.2 (62/68)	98.4 (127/129)	95.9 (189/197)	197

**2. Sensitivity (Detection limit)**

The minimum detection limit is  $5 \times 10^2$  viral particle/test (using 100 µL sample of  $5 \times 10^3$  viral particle/mL for one test).

**3. Reactivity**

Capilia Adeno Neo is responsive to Adenovirus type 1 - 8, 11, 19, 37, 53 and 54.

**4. Cross reactivity**

No cross-reactivity was found in all the viruses and bacteria listed below.

1) Bacteria

- |   |                                       |
|---|---------------------------------------|
| <i>Acinetobacter baumannii</i>                  | <i>Bacillus cereus</i>                |
| <i>Branhamella catarrhalis</i>                  | <i>Bordetella pertussis</i>           |
| <i>Citrobacter freundii</i>                     | <i>Capnocytophaga ochracea</i>        |
| <i>Enterococcus faecalis</i>                    | <i>Enterobacter cloacae</i>           |
| <i>Fusobacterium nucleatum</i>                  | <i>Eikenella corrodens</i>            |
| <i>Haemophilus influenzae</i>                   | <i>Gardnerella vaginalis</i>          |
| <i>Kingella kingae</i>                          | <i>Haemophilus parainfluenzae</i>     |
| <i>Lactobacillus casei</i>                      | <i>Klebsiella oxytoca</i>             |
| <i>Mycobacterium avium</i>                      | <i>Mycobacterium abscessus</i>        |
| <i>Mycobacterium tuberculosis</i>               | <i>Mycobacterium intracellulare</i>   |
| <i>Nocardia asteroides</i>                      | <i>Neisseria meningitidis</i>         |
| <i>Peptostreptococcus anaerobius</i>            | <i>Pasteurella multocida</i>          |
| <i>Prevotella intermedia</i>                    | <i>Porphyromonas asaccharolyticus</i> |
| <i>Salmonella choleraesuis</i> (sub, Minnesota) | <i>Prevotella melaninogenica</i>      |
| <i>Staphylococcus aureus</i>                    | <i>Serratia marcescens</i>            |
| <i>Streptococcus bovis</i> (II Group D)         | <i>Staphylococcus epidermidis</i>     |
| <i>Streptococcus</i> Group B                    | <i>Streptococcus</i> Group A          |
| <i>Streptococcus</i> Group F                    | <i>Streptococcus</i> Group C          |
| <i>Streptococcus milleri</i>                    | <i>Streptococcus</i> Group G          |
| <i>Streptococcus oralis</i>                     | <i>Streptococcus mutans</i>           |
| <i>Streptococcus sanguis</i>                    | <i>Streptococcus pneumoniae</i>       |
| <i>Chlamydophila psittaci</i>                   | <i>Chlamydophila pneumoniae</i>       |

2) Viruses

- |                                 |                                 |
|---------------------------------|---------------------------------|
| Influenza virus A (H1N1)        | Influenza virus A (H3N2)        |
| Influenza virus B               | Influenza virus C               |
| Parainfluenza virus Type 1      | Parainfluenza virus Type 2      |
| Parainfluenza virus Type 3      | Parainfluenza virus Type 4      |
| Respiratory syncytial virus (A) | Respiratory syncytial virus (B) |
| Rhinovirus Type 2               | Coxsackievirus Type A9          |
| Coxsackievirus Type A16         | Coxsackievirus Type B1          |
| Coxsackievirus Type B2          | Coxsackievirus Type B3          |
| Coxsackievirus Type B4          | Coxsackievirus Type B5          |
| Coxsackievirus Type B6          | Echovirus Type 4                |
| Echovirus Type 6                | Echovirus Type 9                |
| Echovirus Type 11               | Echovirus Type 14               |
| Echovirus Type 16               | Cytomegalovirus                 |
| Human Metapneumovirus           |                                 |

**INTERFERING SUBSTANCES**

The following substances were found to have no effect on the results at the concentrations indicated.

Whole blood (0.25%), acetylsalicylic acid (20 mg/mL), acetaminophen (10 mg/mL), ibuprofen (11.25 mg/mL), ambroxol hydrochloride (375 ng/mL), oxymetazoline hydrochloride (100 ng/mL), oseltamivir (7.5 mg/mL), L-carbocysteine (12.5 mg/mL), disodium cromoglycate (5 mg/mL), zanamivir (500 ng/mL), salicylamide (6.75 mg/mL), cyproheptadine hydrochloride hydrate (200 ng/mL), cefixime (2.5 mg/mL), dextromethorphan hydrochloride (10 mg/mL), naphazoline nitrate (125 ng/mL), nifedipine (1 mg/mL), fluticasone propionate (127.5 ng/mL), chlorpheniramine maleate (5 mg/mL), levofloxacin (2.5 mg/mL), and loxoprofen sodium (3 mg/mL)

**REFERENCES**

- Shimizu H, Ishimaru Y, Fujimoto T. Evaluation of Immunochromatographic Detection Kit Using Adenovirus Pt-Au Colloid. *J. J. A. Inf. D.* 2009;83:64-65.
- TAKASAKI Y, SHINDO S, YAMASHITA Y, YOKOYAMA T, SHIBAO K. Evaluation of New Rapid Test Kit "ImmunoAce Adeno" Using Pt-Au Colloid. *Jpn J Clin Exp Med.* 2009;86(5):672-675.
- TAKASAKI Y et al. Comparison Study of Nasal Swab and Nasal Aspirate Using Immunochromatographic Adenovirus Detection Kit. *Jpn J Clin Exp Med.* 2010;87:1168-1171.

**INQUIRES**



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**GLOSSARY OF SYMBOLS**

	CE Marking (European directive 98/79/EC on <i>in vitro</i> diagnostic medical devices)		Authorized representative in the European Community
	<i>In vitro</i> diagnostic medical device		Do not reuse
	Temperature limitation		Manufacturer/Manufactured by
	Use by YYYY-MM		Consult instructions for use
	Batch code		Caution, consult accompanying documents.
	Catalog number		Keep away from sunlight
	Contents sufficient for <n> tests		Fragile, handle with care
	Open here		